

# INTERNATIONAL JOURNAL OF PHARMACY & LIFE SCIENCES

# Cytokines & their physiologic and pharmacologic functions in

inflammation: A review

P. Zuber Shaikh

Sinhgad College of Pharmacy, Pune, (MH) - India

#### Abstract

Inflammation is mediated by a variety of soluble factors, including a group of secreted polypeptides known as cytokines. Inflammatory cytokines can be divided into two groups: those involved in acute inflammation and those responsible for chronic inflammation. This review describes the role played in acute inflammation by IL-1, TNF- $\alpha$ , IL- 6, IL-11, IL-8 and other chemokines, G-CSF, and GM-CSF. It also describes the involvement of cytokines in chronic inflammation. This latter group can be subdivided into cytokines mediating humoral responses such as IL-4, IL-5, IL-6, IL-7, and IL-13, and those mediating cellular responses such as IL-1, IL-2, IL-3, IL-4, IL-7, IL-9, IL-10, IL-12, interferons, transforming growth factor- $\beta$ , and tumor necrosis factor  $\alpha$  and  $\beta$ . Some  $\alpha$  cytokines, such as IL-1, significantly contribute to both acute and chronic inflammation. This review also summarizes features of the cellsurface receptors that mediate the inflammatory effects of the described cytokines. The anti-inflammatory cytokines are a series of immunoregulatory molecules that control the proinflammatory cytokine response. Cytokines act in concert with specific cytokine inhibitors and soluble cytokine receptors to regulate the human immune response. Their physiologic role in inflammation and pathologic role in systemic inflammatory states are increasingly recognized. Major anti-inflammatory cytokines include interleukin (IL)-1 receptor antagonist, IL-4, IL-6, IL-10, IL-11, and IL-13. Specific cytokine receptors for IL-1, tumor necrosis factor-  $\alpha$ , and IL-18 also function as proinflammatory cytokine inhibitors. The nature of anti-inflammatory cytokines and soluble cytokine receptors is the focus of this review.

Key-Words: anti-inflammatory cytokines, Cytokines, Inflammation, Sepsis, Septic shock

# Introduction

Inflammation, the response of tissue to injury, is characterized in the acute phase by increased blood flow & vascular permeability along with the accumulation of fluid, leukocytes, & inflammatory mediators such as cytokines. In the subacute / chronic phase (hereafter reffered to as the chronic phase), it is charactrised by the development of specific humoral & cellular immune responses to the pathogens present at the site of tissue injury.<sup>1</sup> The human immune response is regulated by a highly complex and intricate network of control elements. Under physiologic conditions, these cytokine inhibitors serve as immunomodulatory elements that limit the potentially injurious effects of sustained or excess inflammatory reactions.

\* Corresponding Author: E-mail:zuby.shaikh56@gmail.com, zuby.shaikh20@gmail.com Under pathologic conditions, these anti-inflammatory mediators may either (1) provide insufficient control over proinflammatory activities in immune-mediated diseases or (2) overcompensate and inhibit the immune response, rendering the host at risk from systemic infection.<sup>2,3</sup> A dynamic and ever-shifting balance exists between proinflammatory cytokines and anti-inflammatory components of the human immune system.

The regulation of inflammation by these cytokines and cytokine inhibitors is complicated by the fact that the immune system has redundant pathwayswith multiple elements having similar physiologic effects. Furthermore, with the potential exception of interleukin (IL)-1 receptor antagonist (IL-1ra), all the antiinflammatory cytokines have at least some proinflammatory properties as well. The net effect of any cytokine is dependent on the timing of cytokine release, the local milieu in which it acts, the presence of competing or synergistic elements, cytokine receptor density, and tissue responsiveness to each cytokine.<sup>4</sup>

This is what makes the study of cytokine biology so fascinating (and so frustrating as well!). Perturbations of this regulatory network of cytokines by genetic, environmental, or microbial elements may have highly deleterious consequences.<sup>5-9</sup> These inhibitory cytokines have already proven to be efficacious in a variety of clinical conditions marked by excess inflammation. Their potential therapeutic use in numerous other inflammatory states will also be described. The functional definition of an anti-inflammatory cytokine in this review is the ability of the cytokine to inhibit the synthesis of IL-1, tumor necrosis factor (TNF), and other major proinflammatory cytokines.

# Cytokines involved in acute inflammation:

Several cytokines play key roles in mediating acute inflammatory reactions, namely IL-1, TNF- $\alpha$ , IL-6, IL-11, IL-8 & other chemokines, G-CSF, and GM-CSF. Of these, IL-1( $\alpha$  and  $\beta$ ) & TNF are extremely potent inflammatory molecules: they are the primary cytokines that mediate acute inflammation induced in animals by intradermal injection of bacterial lipopolysaccharide.

#### Interleukin-1:

Their main cellular sources are mononuclear phagocytes, fibroblasts, keratinocytes, and T and B synonyms--endogenous lymphocytes. Previous pyrogen (EP), mononuclear cell factor. and lymphocyte-activating factor (LAF)--emphasize the role of IL-1 in inflammation. Both IL-1 $\alpha$  and IL-1 $\beta$ can trigger fever by enhancing prostaglandin  $E_2$  (PGE<sub>2</sub>) synthesis by the vascular endothelium of the hypothalamus and can stimulate T cell proliferation. In addition, IL-1 elicits the release of histamine from mast cells at the site of inflammation. Histamine then triggers early vasodilation and increase of vascular permeability. The pro-inflammatory effects of IL-1 can be inhibited by IL-1 receptor antagonist (IL-1Ra), originally referred to as IL-1 inhibitor. IL-1Ra is produced by immune complex- or IL-4-stimulated macrophages and by TNF- or GM-CSF-stimulated neutrophils. It bears approximately 20-25% homology at the amino acid level to  $IL-1\alpha$  and  $IL-1\beta$ . IL-1Ra inhibits IL-1 action by competing with IL-1 for binding to the IL-1 receptor (IL-1R).<sup>10,11</sup>

# IL-1ra

IL-1ra is produced by monocytes and macrophages and is released into the systemic circulation in >100-fold excess than either IL-1a or IL-1b after lipopolysaccharide (LPS) stimulation in human volunteers. The synthesis of IL-1ra and IL-1 $\beta$  are differentially regulated at their own promoter sites. Although bacterial LPS stimulates the synthesis of both

# [Shaikh, 2(11): Nov., 2011] ISSN: 0976-7126

IL-1ß and IL-1ra, other stimuli cause differential release of IL-1ra and IL-1β. The anti-inflammatory cytokines IL-4, IL-6, IL-10, and IL-13 inhibit the synthesis of IL-1b, yet they stimulate the synthesis of IL-1ra. There is at least one important polymorphism in the genetic regulation of IL-1ra synthesis in human populations.<sup>12</sup> DNA polymorphisms at this site may determine the synthetic rate of IL-1ra and alter the host response to inflammatory stimuli. Excess IL-1ra synthesis in relationship to IL-1 $\alpha$  or IL-1 $\beta$  synthesis has been shown to increase susceptibility to diverse human pathogens such as Lyme arthritis, tuberculosis, and a variety of other infectious diseases. Conversely, inadequate local IL-1ra synthesis in the lung may predispose to severe acute lung injury and result in excess lethality in ARDS. Because IL-1 is such a prominent proinflammatory cytokine in a multitude of systemic inflammatory states, IL-1ra has been extensively studied in clinical trials as a specific IL-1 inhibitor. Despite convincing evidence that IL-1 plays an important role in the pathogenesis of bacterial sepsis, the results of IL-1ra therapy in large phase III clinical trials for severe sepsis have been disappointing. Nonetheless, IL-1ra continues to be a promising new treatment for the management of patients with refractory forms of rheumatoid arthritis.<sup>16</sup>

# Interleukin-2:

IL-2 is a glycoprotein originally known as T cell growth factor (TCGF). It is secreted mainly by activated T helper cells. It acts as a growth factor/activator for T cells, NK cells, and B cells and promotes the development of lymphokine-activated killer (LAK) cells. It therefore plays a critical role in regulating both cellular and humoral chronic inflammatory responses. Binding of IL-2 to the IL-2 receptor on T lymphocytes leads to cell proliferation, increased lymphokine secretion, and enhanced expression of class II MHC molecules.

#### Interleukin-3:

IL-3, also called multi-CSF, is produced by activated T cells and mast cells. It stimulates eosinophils and B cell differentiation while it inhibits lymphokine-activated killer (LAK) cell activity<sup>28</sup>. IL-3 shares several biological activities with GM-CSF.<sup>17</sup>

#### Interleukin-4:

IL- 4 is produced by CD4+ ( $T_H$ ) cells, mast cells, and basophils. It induces CD4+ T cells to differentiate into  $T_H2$  cells while suppressing the development of  $T_H1$ cells. It also acts as a B cell, T cell, and mast cell growth factor, it enhances class II MHC expression on B cells, and it promotes immunoglobulin class switching to IgG<sub>1</sub> and IgE. In fact, IL-4 is necessary for

IgE response induction, and its absence also leads to significantly lower levels of IgG1 in T cell-dependent immune responses.<sup>18</sup> The stimulatory effects of IL-4 on IgG1 and IgE production and on MHC class II induction are downregulated by IFN-y, a cytokine whose functions are antagonized by IL-4 and vice versa. IL-4 also stimulates collagen <sup>19</sup> and IL-6 production <sup>20</sup> by human dermal fibroblasts, and may thus play a role in the pathogenesis of fibrotic diseases such as systemic sclerosis. In rheumatoid arthritis, on the other hand, IL-4 appears to exhibit some antiinflammatory properties by inhibiting the production of several proinflammatory cytokines such as IL-1, IL-6, IL-8, and TNF- $\alpha$ , by synovial membranes of rheumatoid arthritis patients.<sup>21</sup> IL-4 is a highly pleiotropic cytokine that is able to influence Th cell differentiation. Early secretion of IL-4 leads to polarization of Th cell differentiation toward Th2-like cells. Th2-type cells secrete their own IL-4, and subsequent autocrine production of IL-4 supports cell proliferation. The Th2- cell secretion of IL-4 and IL-10 leads to the suppression of Th1 responses by downregulating the production of macrophage-derived IL-12 and inhibiting the differentiation of Th1-type cells. IL-4 drives Th2 responses, mediates the recruitment and activation of mast cells, and stimulates the production of IgE antibodies via the differentiation of B cells into IgE-secreting cells.<sup>22,23</sup> IL-4 has marked inhibitory effects on the expression and release of the proinflammatory cytokines. It is able to block or suppress the monocyte-derived cytokines, including IL-1, TNF- $\alpha$ , IL-6, IL-8, and macrophage inflammatory protein (MIP)-1 $\alpha$ .<sup>24,25</sup> It has also been shown to suppress macrophage cytotoxic activity, parasite killing, and macrophage-derived nitric oxide production.<sup>26</sup> In contrast to its inhibitory effects on the production of proinflammatory cytokines, it stimulates the synthesis of the cytokine inhibitor IL-1ra.<sup>27</sup> The immunologic effects of IL-4 in the presence of bacterial infection are complex and incompletely understood. IL-4 has been shown to enhance clearance of Pseudomonas aeruginosa from lung tissue in experimental models of Gram-negative bacterial pneumonia.<sup>28</sup> In Gram-positive bacterial infection models, IL-4 has been found to act as a growth factor for Staphylococcus aureus, resulting in systemic infection and increased lethality from bacterial sepsis. <sup>29</sup> IL-4 is able to affect a variety of structural cells. It can potentiate proliferation of vascular endothelium and skin fibroblasts yet decrease proliferation of adult human astrocytes and vascular smooth muscle cells.<sup>30</sup> In addition, IL-4 induces a potent cytotoxic response against tumors.<sup>31,32</sup> IL-4 may act by stabilizing disease

and modifying tumor growth rates in addition to inducing tumor shrinkage and cell death without causing severe side effects, suggesting a possible adjuvant role for IL-4 in the treatment of malignant diseases.

# **Interleukin-5:**

IL-5, also known as B cell growth factor II (BCGFII) and T cell replacing factor (TRF), is produced by CD4+ T helper cells as well as NK cells. IL-5 is involved in eosinophil differentiation and activation and stimulation of immunoglobulin class switching to IgA. Other properties of IL-5 include increased activation of B cell proliferation, and enhancement of T cell cytotoxicity. The combined production of IL-4 and IL-5 by CD4+  $T_{H2}$  cells therefore results in IgE and IgA production and mast cell and eosinophil stimulation.

# **Interleukin-6:**

Previous synonyms of IL-6 illustrate some of its biologic activities. They include interferon- $\beta_2$  (IFN- $\beta_2$ ). hybridoma/plasmacytoma growth factor, hepatocytestimulating factor, B cell stimulatory factor 2 (BSF-2), and B cell differentiation factor (BCDF). IL-6 is produced by a variety of cells including mononuclear phagocytes, T cells, and fibroblasts.<sup>33,34</sup> In addition to the stimulation of acute phase protein synthesis by the liver, IL-6 acts as a growth factor for mature B cells and induces their final maturation into antibodyproducing plasma cells. It is involved in T cell activation and differentiation, and participates in the induction of IL-2 and IL-2 receptor expression. Some of the regulatory effects of IL-6 involve inhibition of TNF production, providing negative feedback for limiting the acute inflammatory response. Upregulation of IL-6 production has been observed in a variety of chronic inflammatory and autoimmune disorders such as thyroiditis, type I diabetes, rheumatoid arthritis<sup>35,36</sup> sclerosis<sup>37</sup>, mesangial proliferative systemic glomerulonephritis and psoriasis, and neoplasms such as cardiac myxoma, renal cell carcinoma, multiple myeloma, lymphoma, and leukemia. IL-6 has long been regarded as a proinflammatory cytokine induced by LPS along with TNF-a and IL-1. IL-6 is often used as a marker for systemic activation of proinflammatory cytokines.<sup>38</sup> Like many other cytokines, IL-6 has both proinflammatory and anti-inflammatory properties. Although IL-6 is a potent inducer of the acute-phase protein response, it has anti-inflammatory properties as well.<sup>39</sup> Inasmuch as these peptide molecules use a common cellular receptor, they share many of the physiologic features attributable to IL-6. IL-6 downregulates the synthesis of IL-1 and TNF.40,41 IL-6 attenuates the synthesis of the proinflammatory

cytokines while having little effect on the synthesis of anti-inflammatory cytokines such as IL-10 and transforming growth factor- $\beta$  (TGF- $\beta$ ). IL-6 induces the synthesis of glucocorticoids<sup>42</sup> and promotes the synthesis of IL- 1ra and soluble TNF receptor release in human volunteers.<sup>43</sup> At the same time, IL-6 inhibits the production of proinflammatory cytokines such as GM-CSF, IFN- $\gamma$ , and MIP-2. The net result of these immunologic effects place IL-6 among the antiinflammatory cytokine group.

# **Interleukin-7:**

IL-7 is a cytokine known as a pre-B cell growth factor, is a bone marrow and thymic stromal cell product. It stimulates the development of pre-B and pre-T cells and acts as a growth factor for B cells, T cells, and early thymocytes.44

#### **Interleukin-8/chemokines:**

IL-8 and other low molecular weight chemokines (e.g. platelet factor 4, macrophage inflammatory protein (MIP)-1 $\alpha$  and  $\beta$ , MIP-2, monocyte chemoattractant protein-1 (MCP- 1/JE), RANTES) belong to a chemotactic cytokine family and are responsible for the chemotactic migration and activation of neutrophils and other cell types (such as monocytes, lymphocytes, basophils, and eosinophils) at sites of inflammation. 45,46 The two subsets of the chemokine family, "CXC" (or  $\alpha$ ), "C-C" (or  $\beta$ ) are divided based on presence or absence of an amino acid between the first two of four conserved cysteines. A recent third subset, "C", has only two cysteines and to date only one member, IL-16, has been identified.<sup>47</sup> Chemokines have been implicated in inflammatory conditions from acute neutrophil-mediated conditions such as acute respiratory distress syndrome to allergic asthma, arthritis, psoriasis, and chronic inflammatory disorders. To date, at least 27 chemokines have been described. The product of many cell types, including mononuclear phagocytes, antigen-activated T cells, endothelial and epithelial cells, and even neutrophils, IL-8 was previously known as neutrophil chemotactic factor (NCF) and neutrophil activating protein (NAP-1).<sup>48</sup> It is the most thoroughly studied chemokine and therefore serves as a prototype for discussing the biologic properties of this rapidly growing family of inflammatory mediators. Its main inflammatory impact lies in its chemotactic effects on neutrophils and its ability to stimulate granulocyte activity. In addition, IL-8, IL-1, and TNF are involved in neutrophil recruitment by upregulating cell-surface adhesion molecule expression (such as endothelial leukocyte adhesion molecule, ELAM-1, and intracellular adhesion molecule, ICAM-1), thereby enhancing neutrophil adherence to endothelial cells and facilitating their diapedesis through vessel walls. Thus, IL-8 mediates the recruitment and activation of neutrophils in inflamed tissue.<sup>49</sup> IL-8 can be detected in synovial fluid from patients with various inflammatory rheumatic diseases,<sup>50</sup> and mucosal levels of IL-8 are elevated in patients with active ulcerative colitis.<sup>51</sup> Other members of this cytokine family, such as NAP-2, RANTES, MCP-1, MCP-2, MCP-3, platelet factor 4, MIP-1 $\alpha/\beta$ , and MIP-2, are also likely to play important roles in acute inflammation via their shared effects on cell migration. MCP-1 is a chemokine identified in supernatants of blood mononuclear cells. Its production in monocytes is enhanced by inflammatory cytokines. MIP-1a and MIP-1B

induce monocyte and T lymphocyte migration. MIP- $1\alpha$ , MCP-1, and MIP-2 have been implicated in the pathogenesis of rheumatoid arthritis where they are believed to recruit mononuclear cells into the inflamed regions of the synovium.<sup>52</sup> Several other members of the IL-8/chemokine family have been identified but their biologic effects are as yet poorly defined. Two recently identified chemokines, eotaxin and IL-16, have some unique properties and are described below.

# **Interleukin-9:**

IL-9 is another cytokine produced by CD4+ T helper  $(T_{H}2)$  cells as well as some B lymphomas first described in the mouse, IL-9 was known as mast cell growth-enhancing activity (MEA) and murine Tcell growth factor P40.53 Its production is IL-4 and IL-10, and thus IL-2-dependent. IL-9 is regulatory in nature in that it inhibits lymphokine production by IFN-yproducing CD4+T cells and enhances the growth of CD8+T cells.<sup>54</sup> In addition, IL-9 promotes the production of immunoglobulins by B cells and the proliferation of mast cells.55

# **Interleukin-10:**

IL-10 is also referred to as B cell-derived T cell growth factor and cytokine synthesis inhibitory factor (CSIF) because it inhibits IFN- $\gamma$  production by activated T cells. IL-10 is produced by a variety of cell types, including CD4+ T cells, activated CD8+ T cells, and activated B cells.<sup>56</sup> Its effects include reduction of antigen-specific T cell proliferation, inhibition of IL- 2induced IFN- $\gamma$  production by NK cells, and inhibition of IL-4 and IFN- $\gamma$  induced MHC class II expression on monocytes.<sup>56</sup> Since IL-10 can be produced by  $T_{H2}$  cells and inhibits  $T_{H1}$  function by preventing  $T_{\rm H}1$  cytokine production (such as IFN- $\gamma$ ), IL-10 is considered a T cell cross-regulatory factor and has thus been referred to as an "anticytokine".<sup>57</sup> IL-10 also acts as a co-differentiation factor for cytotoxic T cells and a co-factor for T cell growth. Human IL-10

(hIL-10) shares 84% identity at the amino acid level with a homolog, viral IL-10 (vIL-10), which is encoded by the Epstein-Barr virus.<sup>58</sup> vIL-10 shares with hIL-10 inhibitory effects on cytokine production and stimulatory effects on B cell growth.<sup>59</sup> IL-10 is the most important anti-inflammatory cytokine found within the human immune response. It is a potent inhibitor of Th1 cytokines, including both IL-2 and IFN-γ. This activity accounts for its initial designation as cytokine synthesis inhibition factor.<sup>60,61</sup> In addition to its activity as a Th2 lymphocyte cytokine, IL-10 is also a potent deactivator of monocyte/macrophage proinflammatory cytokine synthesis.<sup>62,63</sup> IL-10 is primarily synthesized by CD41 Th2 cells, monocytes, and B cells. After engaging its high-affinity 110-kd cellular receptor, IL-10 inhibits monocyte/macrophagederived TNF-a, IL-1, IL-6, IL-8, IL-12, granulocyte colony-stimulating factor, MIP-1a, and MIP-2a.<sup>64-65</sup> IL-10 inhibits cell surface expression of major histocompatibility complex class II molecules, B7 accessory molecules, and the LPS recognition and signaling molecule CD14. It also inhibits cytokine production by neutrophils and natural killer cells. IL-10 inhibits nuclear factor kB  $(NF-\kappa\beta)$  nuclear translocation after LPS stimulation and promotes degradation of messenger **RNA** for the proinflammatory cytokines. In addition to these activities, IL-10 attenuates surface expression of TNF receptors and promotes the shedding of TNF receptors into the systemic circulation.<sup>66-67</sup>IL-10 is readily measurable in the circulation in patients with systemic illnesses and a variety of inflammatory states.<sup>68,69</sup> IL-10 is present in sufficient concentrations to have a physiologic impact on host responses to systemic inflammation. It has been determined that patients who preferentially express high levels of IL-10 and reduced levels of TNF- $\alpha$  are more likely to die from meningococcemia<sup>70,71</sup> and a variety of other community acquired infections.<sup>72</sup> Physiologically inadequate IL-10 responses after systemic injury may have detrimental consequences as well. Low lung concentrations of IL-10 in patients with acute lung injury indicate that ARDS is more likely to develop. The administration of IL-10 in experimental animal models of endotoxemia improves survival. Human volunteers given IL-10 after endotoxin challenge suffer fewer systemic symptoms, neutrophil responses, and cytokine production than placebo-treated control subjects.<sup>73</sup> Moreover, mice who have genetic deletions of the IL-10 gene are more susceptible to endotoxin-induced shock than normal mice.<sup>74</sup> IL-10 generally protects the host from systemic inflammation after toxin-induced injury, but renders the host susceptible to lethality from overwhelming

# [Shaikh, 2(11): Nov., 2011] ISSN: 0976-7126

infection in a variety of experimental studies.<sup>75,76</sup> This observation should be kept in mind when administering anti-inflammatory cytokines in clinical medicine. The IL-10 knockout mouse spontaneously develops a chronic inflammatory enteritis that mimics inflammatory bowel disease in humans.<sup>77-80</sup> This indicates that endogenous concentrations of IL-10 are important in limiting the inflammatory response to gut-associated bacteria. For this reason, IL-10 is in clinical trials as an anti-inflammatory therapy for inflammatory bowel disease among other potential indications.

# Interleukin-11:

IL-11is produced by bone marrow stromal cells and by some fibroblasts. It is a functional homologue of IL-6 and can replace IL-6 for the proliferation of certain plasmacytoma cell lines and in the induction of acute phase protein secretion in the liver.<sup>81</sup> Additional IL-11 activities include stimulation of T cell-dependent B cell immunoglobulin secretion, increased platelet production, and induction of IL-6 expression by CD4+ T cells. IL-11 shares many properties of IL-6, including the common use of the gp130 receptor ligand complex as a signal transduction pathway. IL-11 binds to its own unique receptor and then complexes with gp130 cell membranes of target cells.<sup>82</sup> IL-11 was initially described as a hematopoietic growth factor with particular activity in the stimulation of thrombopoiesis. IL-11 has recently been approved for clinical use as a platelet restorative agent after chemotherapy-induced bone marrow suppression.<sup>83</sup> It has become clear that IL-11 has important immunoregulatory activities separate from its hematopoietic growth factor potential. IL-11 has been shown to attenuate IL-1 and TNF synthesis from macrophages by up-regulating inhibitory NF-kB (inhibitory NF- $\kappa\beta$ ) synthesis in monocyte/macrophage cell lines. Inhibitory NF-kB prevents NF-kB from translocating to the nucleus where NF-kB functions as a transcriptional activator for the proinflammatory cytokines.<sup>84</sup> IL-11 has also been shown to inhibit the synthesis of IFN- $\gamma$  and IL-2 by CD41 T cells. IL-11 functions as a Th2-type cytokine, with induction of IL-4 and inhibition of Th1-type cytokines.<sup>85</sup> IL-11 does not induce the synthesis of IL-10 or TGF-β. This indicates that IL-11 is a direct inhibitor of Th1 lymphocytes and does not act indirectly through induction of IL-10. IL-11 is rarely measurable in the systemic circulation but has been detected and is physiologically active in localized areas of inflammation, such as inflammatory arthritis or inflammatory bowel disease.86 IL-11 is currently in clinical trials as an immunomodulator for a number of potential clinical indications.

# Interleukin-12:

IL-12, previously known as natural killer cell stimulatory factor (NKSF) and cytotoxic lymphocyte maturation factor (CLMF), was originally isolated from Epstein-Barr virus transformed B cells. Its biological activities include enhancement of cytotoxic T cells and lymphokine activated killer (LAK) cell generation and activation, increased natural killer (NK) cell cytotoxicity, induction of activated T cell and NK cell proliferation, induction of IFN-y production by NK cells and T cells, and inhibition of IgE synthesis by IL-4-stimulated lymphocytes via IFN- $\gamma$ -dependent and independent mechanisms.<sup>87,88</sup> IL-12 is secreted by activated B cells, macrophages, and other antigenpresenting cells (APCs), but its production is inhibited by IL-4 and IL-10. In addition, the stimulatory effect of IL-12 on  $T_{\rm H}$  development is antagonized by IL-4, a cytokine which promotes  $T_H^2$  cell development. Therefore, IL-12 plays an important role in cellmediated inflammation and also contributes to the regulation of immunoglobulin production.

# Interleukin-13:

IL-13 exhibits anti-inflammatory activities by inhibiting the production of inflammatory cytokines, such as IL-1β, TNF-α, IL-8, and IL-6, by human peripheral blood monocytes induced with lipopolysaccharide. Inhibition of inflammatory cytokine production is also a characteristic of two other cytokines produced by T<sub>H</sub>2 lymphocytes, namely IL-4 and IL-10. In addition, IL- 13 enhances monocyte and В lymphocyte differentiation and proliferation, increases CD23 expression, and induces IgG4 and IgE class switching.<sup>89</sup> IL-13, a potent in vitro modulator of human monocytes and B-cell function, is secreted by activated T lymphocytes.<sup>90,91</sup> IL-13and IL-4 share a common cellular receptor (IL-4 type 1 receptor), and this accounts for many of the similarities between these two anti-inflammatory cytokines.<sup>92</sup> IL-4 and IL-13 share only 20% to 25% primary amino acid homology, but the major a-helical regions that are essential for their activity are highly homologous. The principal functional difference between IL-4 and IL-13 lies in their effects on T cells. IL-4 is a dominant mediator of Th2 cell differentiation, proliferation, and activity, whereas IL-13 has minimal effects on T-cell function. IL-13 can down-regulate the production of TNF, IL-1, IL-8, and MIP-1 $\alpha$  by monocytes and has profound effects on expression of surface molecules on both monocytes and macrophages. IL-13 upregulates cell surface expression of  $b_2$  integrins and major histocompatibility complex (MHC) class II antigens and down-regulates CD14 and Fcy receptor expression.

IL-13 inhibits NF-kB activation in macrophages and protects against LPS-induced lethality in animal models.<sup>93,94</sup> IL-13 suppresses lung inflammatory injury after the deposition of IgG immune complexes.<sup>95-97</sup> Exogenous administration of anti-inflammatory cytokines into the lungs of rats after IgG immune complex deposition reveals that the greatest inhibitory activity is observed by IL-13 and IL-10, followed by IL-4 and IL-6. The potential role of IL-13 in clinical medicine remains to be defined.<sup>98</sup>

# Interleukin-14:

A product of malignant B and T cells as well as normal T cells, B-cell growth factor (BCGF). Like IL-4, IL-14 has been shown to induce B cell proliferation. However, IL-14 inhibits immunoglobulin secretion. It has been suggested to play an important role in the aggressive form of B-cell type non-Hodgkin's lymphoma.

# Interleukin-15:

IL-15 is a cytokine of a T cell stimulatory activity produced by activated monocytes, epithelial cells, and fibroblasts. IL-15 shares many biologic properties with IL-2 and mediates its activity via a multi-subunit high affinity receptor comprised of a unique alpha chain and the beta and gamma chains of the IL-2R. IL-15 is produced by a large variety of cells including T lymphocytes and monocytes. It stimulates T lymphocyte and NK cell proliferation. It enhances B cell expansion and immunoglobulin production.<sup>99</sup> It is also a T lymphocyte chemoattractant. IL-15 may be responsible for the recruitment and activation of T lymphocytes in the synovium of patients with rheumatoid arthritis where its levels have been found to be elevated.<sup>100</sup>

# Interleukin-16:

IL-16 was originally identified as a chemotactic factor known as lymphocyte chemoattractant factor or lymphotactin. It is the only member of the "C" family of chemokines. IL-16 is an unusual cytokine in that preformed IL-16 is stored in CD8+ lymphocytes and is secreted upon stimulation with histamine or serotonin.<sup>101</sup> It induces chemotaxis of CD4+ T lymphocytes <sup>102,103</sup> and is believed to initiate T-cell mediated inflammation in asthma.<sup>104</sup>

# Interleukin-17:

IL-17 is a product of activated T lymphocytes and its biologic activities include stimulation of IL-6 and IL-8 production and enhanced ICAM-1 expression on human foreskin fibroblasts.<sup>105</sup>

# Tumor necrosis factor:

Tumor necrosis factors-(TNF)  $\alpha$  and  $\beta$  are cytokines that bind to common receptors on the surface of target cells and exhibit several common biological activities. TNF- $\alpha$ , or cachectin, exists as a trimer <sup>131</sup> and is one of the products of activated macrophages/monocytes, fibroblasts, mast cells, and some T and natural killer (NK) cells.<sup>106,107</sup> TNF-α and IL-1 share several proinflammatory properties. Like IL-1, TNF- α can induce fever, either directly via stimulation of PGE<sub>2</sub> synthesis by the vascular endothelium of the hypothalamus, or indirectly by inducing release of IL-1. Both cytokines can stimulate the production of collagenase and PGE<sub>2</sub> by synovial cells and thus are believed to contribute to joint damage in inflammatory conditions such as rheumatoid arthritis. TNF- $\alpha$  also shares an important inflammatory property with IL-6 and IL-11, i.e. the induction of acute phase reactant protein production by the liver. TNF- $\alpha$  and IL-1 further exert secondary inflammatory effects by stimulating IL-6 synthesis in several cell types. IL-6 then mediates its own effects and those of TNF-  $\alpha$  and IL-1 in inducing fever and the acute phase response, thereby perpetuating the inflammatory response through a cascade of cytokines with overlapping properties. TNF- $\alpha$ , also known as lymphotoxin, is produced by activated T and B lymphocytes. It binds to the same high affinity receptors as TNF- $\alpha$ . Its properties are similar to those of TNF- $\alpha$  and include the induction of apoptosis (programmed cell death) in many types of transformed, virally infected, and tumor cells, and the stimulation of several PMN effector functions.<sup>108</sup> Although in general the effects of cytokines are exerted locally at the site of their production (autocrine and paracrine), TNF- a and TNF- $\beta$ , as well as IL-1 and IL-6, have major systemic (endocrine) effects when either produced acutely in large amounts, as in the case of bacterial sepsis, or chronically in lesser amounts, as in the case of chronic infections. During sepsis with Gram negative organisms, lipopolysaccharides (endotoxin) released from bacteria trigger the widespread production of TNF- $\alpha$  (and subsequently IL-1 and IL-6) by macrophages. The systemic release of these cytokines has been shown to be responsible for the fever and hypotension that characterize septic shock. In an analogous fashion, the production of large amounts by T lymphocytes in response to of TNF-α "superantigens" such as staphylococcal toxic shock syndrome toxin and enterotoxins are responsible for many of the systemic manifestations (fever, hypotension) of infections with toxin-producing Gram positive organisms.<sup>109,110</sup> In addition, the chronic production of TNF is believed to be responsible for the

metabolic alterations which result in the cachexia associated with chronic parasitic infections and some cancers.

# **Eotaxin:**

Eotaxin is a specific chemoattractant for eosinophils. It is produced by cytokine-stimulated epithelial and endothelial cells as well as IL-3- stimulated eosinophils. Eotaxin is implicated in inflammatory bowel disease where its mRNA levels are markedly elevated, especially in ulcerative colitis.<sup>111</sup>

# **Colony stimulating factors:**

Colony stimulating factors (CSF) are named according to the target cell type whose colony formation in soft agar cultures of bone marrow they induce.<sup>112</sup> Of the CSF's, granulocyte-CSF (G-CSF) and granulocyte macrophage-CSF (GM-CSF) participate in acute inflammation. G-CSF was cloned in 1986 and its gene was mapped to chromosome 17.<sup>113</sup> Monocytes, T cells, fibroblasts and endothelial cells activated by macrophage products such as IL-1 or TNF, can produce G-CSF and GM-CSF. Both CSF's can stimulate neutrophils, while GM-CSF can also activate effector functions of eosinophils and mononuclear phagocytes. An example of the pathophysiologic role of GM-CSF is the airway inflammation accompanying asthma, where the implicated cytokines include IL-3, IL-5, and GM-CSF which perpetuate eosinophil activation and survival. In this scenario, the source of GM-CSF may be the alveolar macrophages which are reported to produce two to threefold higher levels of GM-CSF than control macrophages. Another possible source for all three cytokines are T cells present in the airways. Additional cytokines such IL-4, IL-13 (both stimulatory) and IFN- $\gamma$  (inhibitory) may be involved in the control of IgE synthesis, while IL-1 and TNF- $\alpha$ may contribute to the airway inflammation by upregulation of endothelial adhesion molecule expression.114

# **Transforming grow**th factor - $\beta$ :

The transforming growth factor-  $\beta$  (TGF- $\beta$ ) family of cytokines includes three isoforms, TGF- $\beta$ 1,  $\beta$ 2, and  $\beta$ 3 which are encoded by separate genes yet bind to the same high affinity receptor. It is produced by T cells, platelets, and monocytes. TGF- $\beta$  inhibits T cell and NK cell proliferation and activation and may play an important role in inflammation. At a site of injury, TGF- $\beta$  stored in platelets is released upon degranulation. TGF- $\beta$  then attracts monocytes and other leukocytes to the site, thus participating in the initial step of chronic inflammation. TGF- $\beta$  then positively regulates its own production and the production and deposition of extracellular matrix components as well as the expression of integrins

resulting in enhanced cell adhesion. It also inhibits collagenase production, and if expression is prolonged, it may result in progressive fibrosis analogous to unregulated tissue repair. Conditions in which a role has been suggested include mesangial for TGF-β glomerulonephritis proliferative and diabetic nephropathy in rats, pulmonary fibrosis, and systemic sclerosis. Another example of the role played by TGF- $\beta$  in inflammation is collagen-induced arthritis in rats. In this model, TNF- $\alpha$  and TGF- $\beta$ , when injected into the rat ankle joint, accelerate disease onset.<sup>115</sup> TGF- $\beta$  is synthesized as an inactive precursor and requires activation before exerting its effect. There are three isoforms of TGF- $\beta$  (designated TGF- $\beta_{1-3}$ ) expressed in mammalian species. TGF- $\beta$  is an important regulator of cell proliferation, differentiation, and formulation of the extracellular matrix. In vitro, it inhibits growth of ectodermally derived cells. TGF- $\beta$  induces squamous cell differentiation of human bronchial epithelial cells.<sup>116</sup> TGF- $\beta$  has been shown to inhibit alveolar type II cell proliferation and to decrease the expression of surfactant protein A in human lung explant cultures and in a human lung adenocarcinoma cell line. TGF-B appears to contribute to the fibroproliferative phase of acute lung injury from a variety of injurious agents. It plays a role in regulating the extracellular matrix by decreasing degradation of matrix proteins through a reduction in protease synthesis and an increase in the synthesis of protease inhibitors. Like many cytokines, TGF- $\beta$  has both pro- and anti-inflammatory effects. It functions as a biological switch, antagonizing or modifying the action of other cytokines or growth factors. The presence of other cytokines may modulate the cellular response to TGF- $\beta$ , and the effect may differ depending on the activation state of the cell. **TGF-\beta is capable of converting an active site of** inflammation into one dominated by resolution and repair. TGF-B often exhibits effects with immuneenhancing activity in local tissues and immunesuppressive activity in the systemic circulation. TGF- $\beta$ 1 suppresses the proliferation and differentiation of T cells and B cells and limits IL-2, IFN-y, and TNF production. TGF- $\beta_1$  acts as a monocyte/ macrophage deactivator in a manner similar to IL- 10. However, TGF- $\beta$  is less potent an inhibitor than IL-10 and has little or no effect on IL-1 production. The severe and uncontrolled inflammatory reactions observed in the TGF- $\beta_1$  knockout mouse attests to the physiologic role of TGF-B as an endogenous anti-inflammatory cytokine.117

# Interferons:

The interferons are a group of cytokines originally identified by and named for their anti-viral activity.<sup>118</sup> Type I interferons include IFN-α, an 18-20 kDa product of leukocytes, and IFN-B, a product of fibroblasts. They exhibit anti-viral as well as antiproliferative properties and upregulate MHC class I expression. Type II interferon, immune interferon or IFN- $\gamma$ , is a homodimer produced by activated T cells and NK cells. IFN- $\gamma$  is known to enhance MHC class I and II expression on nucleated cells and to stimulate many of the effector functions of mononuclear phagocytes. While IFN- $\alpha$  and  $-\beta$  bind to a common receptor, IFN- y recognizes a distinct and specific cell surface receptor. IFN- $\gamma$  has been implicated in the pathogenesis of a variety of autoimmune and chronic inflammatory conditions<sup>119</sup> including murine models of systemic lupus erythematosus,<sup>148</sup> Type I diabetes mellitus,<sup>120</sup> adjuvant-induced arthritis,<sup>121</sup> and experimental cerebral malaria.<sup>122</sup> Based on experiments knock-out mice, one of its primary with IFN-γ functions in vivo appears to be the activation of macrophages to kill intracellular pathogens such as Mycobacteria.<sup>123</sup>

# **IFN-** γ**-inducing factor:**

An IFN- $\gamma$ -inducing activity was identified in murine Kupffer cells and activated macrophages and referred to as IFN- $\gamma$ -inducing factor (IGIF).<sup>124</sup> IGIF induces IFN- $\gamma$  production more potently than does IL- 12 and is involved in the development of T<sub>H</sub>1 cells.

# Pharmacologic role of Anti-inflammatory Cytokines and Cytokine Inhibitors:

A complex network of cytokines is generated in response to a systemic immune challenge. Microbial pathogens may actually use components of the cytokine network to their own advantage. A number of DNA viruses synthesize soluble TNF receptor and IL-1 receptors.<sup>126,127</sup> Epstein-Barr virus mediates the synthesis of viral IL-10 in infected human B cells.<sup>128</sup> These viral-induced anticytokine strategies appear to assist the virus in the promotion of viral replication and evasion of host-derived clearance mechanisms. Several bacterial pathogens have the capacity to alter host cell cytokine synthesis, degrade proinflammatory cytokines, or use cytokine receptors as portals of entry for cellular invasion.<sup>129</sup> Administration of inhibitors of proinflammatory cytokines (antibodies, soluble receptors, and anti-inflammatory cytokines) in experimental models generally provides an advantage in systemic toxicity models such as endotoxin challenge studies. However, in localized infection models, inhibitors of the proinflammatory cytokine

system may be detrimental to the host and precipitate overwhelming infection with excess mortality. This is particularly true in the absence of appropriate antimicrobial therapy against the invading microbial The dichotomous nature of pathogen. antiinflammatory cytokine responses in experimental systems is commonly observed in cytokine biology. Inadequate concentrations of anti-inflammatory cytokines result in excess inflammation, yet excess anti-inflammatory cytokine concentrations disrupt clearance mechanisms of microbial pathogens in the host. Nonetheless, these anti-inflammatory agents must be present in far greater concentrations than those of proinflammatory cytokines to inhibit their actions. Systemic concentrations of soluble cytokine inhibitors IL-1ra and IL-10 indicate that they are of sufficient magnitude to at least partially inhibit proinflammatory cytokine action.<sup>130</sup> These results suggest that there may well be a pharmacologic role for anti-inflammatory cytokines and soluble cytokine receptors in the face of systemic inflammation. Recent evidence indicates that individuals differ in their susceptibility to systemic infection and inflammatory states on the basis of their cytokine profiles and genetic background. Patients and first-degree relatives of patients with meningococcemia are more likely to have fatal infections if they have high ratios of IL-10 to TNF- $\alpha$ . Similarly, patients with high ratios of TNF to soluble TNF receptors are at increased risk of having lethal meningococcal infections. These studies make it clear that alterations in cytokine networks can have a significant impact on the human host response to a variety of infectious agents and inflammatory states. Despite complexities inherent in the human immune response, therapeutic intervention with specific cytokine inhibitors or antiinflammatory cytokines has already been shown to have significant clinical benefits.<sup>131</sup> Several of these agents are already approved for clinical use, and others are undergoing extensive clinical trials for a variety of inflammatory disease states. The ability to rapidly assess the state of the human immune response and regulate this response in the presence of a variety of human disease states has been the goal of immunologists for the past century. Advances in human genetics and immunobiology now provide an opportunity to capitalize on recent discoveries in basic immunology and cytokine biology. It is likely that antiinflammatory cytokines and specific cvtokine inhibitors will increasingly find their way into standard clinical practice as we enter the next millennium.

# **Receptor of inflammatory cytokines:**

Cytokines elicit their responses by binding to specific high affinity cell-surface receptors on target cells and initiating a series of intracellular signal transduction pathways. The receptors of several cytokines and growth factors are homologous within their extracellular domains. These receptors have been grouped into families, the largest of which is the hematopoietin receptor superfamily which includes one or multiple chains of the receptors for erythropoietin, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-9, IL-11, IL-12, IL-13, IL-15, v-mpl oncogene, GM-CSF, G-CSF, prolactin, and growth hormone. The receptors in this family share a common structure of four conserved cysteine residues in the amino-terminal portion of the ligand-binding domain, as well as a conserved stretch of amino acids (WSXWS = Trp-Ser-X-Trp-Ser; X representing a nonconserved residue) proximal to the membrane-spanning region. The receptors also share fibronectin type III domains.<sup>132</sup> Of the abovementioned members of the erythropoietin receptor family, one of the best characterized is the IL-2 receptor (IL-2R). It consists of three polypeptide chains: IL-2R $\beta$  (p70) and IL-2R $\gamma$  (p64), which are expressed on resting T cells, and IL- 2Ra (p55; T cell activation antigen or Tac), which is expressed upon T cell activation. Association of these subunits yields a high affinity receptor for IL-2.<sup>133,134</sup> In addition. Tac (IL-2R $\alpha$ ) is shed from cells in a soluble form, but it has low affinity for IL-2. Another member of the erythropoietin receptor family, IL-6 receptor (IL-6R), consists of an 80 kDa ligand-binding molecule and a 130 kDa nonligand binding signal-transducing subunit (gp130). Both molecules exhibit the structures shared members of the by hematopoietin receptor superfamily.<sup>135</sup> Such a bimolecular complex is also described for IL- 3R, IL-5R, and GM-CSFR. One of the biologic consequences of these receptor complexes is that although cytokines bind to specific receptors, some may share common pathways in eliciting the target cell's response as a result of shared receptor components. As an example, IL-6, IL-11, leukemia inhibitory factor (LIF), and oncostatin M recognize different cellular receptors (by virtue of unique ligandbinding subunits), but share the same signaltransducing receptor subunit (gp130) and similar biological activities. These cytokines may therefore exert their effects via common signal transduction pathways. A group of receptors distantly related to the erythropoietin receptor family consists of the receptors for type I ( $\alpha$  and  $\beta$ ) and type II ( $\gamma$ ) interferons.<sup>136</sup> Receptors in this class share a homologous binding domain of about 210 amino acids and four cysteine

pairs divided equally between the amino and carboxy terminal. Another group of related receptors includes the two receptors for TNF, the receptor for nerve growth factor (NGF), a transmembrane protein, FAS (Apo-1 or CD95), involved in the apoptosis of activated T lymphocytes,<sup>137</sup> and CD40, a cell surface receptor important in B cell growth and isotype switching.<sup>138</sup> The TNF receptors are 55 kDa (TR55) and 75 kDa (TR75) proteins that bind TNF- $\alpha$  and  $\beta$ equally. Their extracellular domains share 28% identity. There is growing evidence that the two receptors may mediate different cellular responses to TNF,<sup>139,140</sup> although there may be crosstalk between the receptors, perhaps at the level of the signalling pathways to which they are coupled. The chemokine receptors are members of the G protein-coupled receptor (GPCR) superfamily and include IL-8R-A, an IL-8-specific receptor, IL- 8R-B, a receptor recognized by IL-8, and other chemokines of the CXC subset. Recently, receptors for the CC subset of chemokines have been identified. They include CC-CKR-1, CC-CKR-2, CCCKR- 3, and CC-CKR-4 and CC-CKR-5.141 A recently described receptor, the Duffy blood group antigen receptor for chemokines (DARC), binds both CXC and CC chemokines. In addition, the identification of new 'orphan' chemokine receptors, for which no ligands have been identified, has been reported.<sup>142</sup> Recently, five groups reported that CCCKR- 5 is a co-receptor for certain strains of HIV-1.<sup>143</sup> A 32-bp deletion in *CKR5* is reported to delay progression to AIDS in infected individuals and may be responsible for the antibody-negative status of individuals exposed to HIV-1.144-16-

# Conclusion

In conclusion, cytokines are key modulators of inflammation. They participate in acute and chronic inflammation in a complex network of interactions. Several cytokines exhibit some redundancy in function and share overlapping properties as well as subunits of their cell surface receptors. Better understanding of the pathways regulated by cytokines will allow the identification and/or development of agents for improved modulation of the inflammatory response for the treatment of autoimmune, infectious, and neoplastic diseases.

# References

- 1. J.I. Gallin, I.M. Goldstein, R. Snyderman, editors: Inflammation. Basic principles and clinical correlates, ed 2, New York, Raven Press (1992)
- 2. Munoz C, Carlet J, Fitting C, et al. Dysregulation of in vitro cytokine production

by monocytes during sepsis. J Clin Invest 1991; 88:1747-1754.

- 3. Kasai T, Inada K, Takakuwa T, et al. Antiinflammatory cytokine levels in patients with septic shock. Res Commun Mol Pathol Pharmacol 1997; 98:34-42.
- 4. Dinarello CA. Interleukin-1, interleukin-1 receptors and interleukin-1 receptor antagonist. Int Rev Immunol 1998; 16:457-499
- Do" cke WD, Randow F, Syrbe U, et al. 5. Monocyte deactivation in septic patients: restoration by interferon gamma treatment. Nat Med 1997; 3:678–681
- Westendorp RGJ, Langermans JAM, Huizinga 6. TWJ, et al. Genetic influence on cytokine production in fatal meningococcal disease. Lancet 1997; 349:170–173
- 7. Donnelly SC, Strieter RM, Reid PT, et al. The association between mortality rates and decreased concentrations of interleukin-10 and interleukin-1 receptor antagonist in the lung fields of patients with the adult respiratory distress syndrome. Ann Intern Med 1996; 125:191-196
- Opal SM, Cross AS, Jhung J, et al. Potential 8. hazards of combination immunotherapy in the treatment of experimental sepsis. J Infect Dis 1996; 173:1415-1421
- van de Poll T, Marchant A, van Deventer SJH. 9. The role of interleukin-10 in the pathogenesis of bacterial infection. Clin Microbiol Infect 1997; 3:605-607
- 10. Mosmann TR, Cherwinski H, Bond MW, et al. Two types of murine helper T-cell clone: I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol 1986; 136:2348-2357
- 11. Kelso A. Th1 and Th2 subsets: paradigms lost? Immunol Today 1995; 16:374–379
- 12. C.A. Dinarello: Reduction of inflammation by decreasing production of interleukin-1 or by specific receptor antagonism. Int J Tiss Reac 14, 65-75 (1992)
- 13. C.A. Dinarello: Interleukin-1. In: The cytokine handbook. Ed: Thomson A., Academic Press, San Diego, CA (1994)
- 14. Fang XM, Schro" der S, Hoeft A, et al. Comparison of two polymorphisms of the interleukin-1 gene family: interleukin- 1 receptor antagonist polymorphism contributes to susceptibility to severe sepsis. Crit Care Med 1999; 27:1330-1334

- 15. Russel DA, Tucker KK, Khinookoswong N, et al. Combined inhibition of interleukin-1 in tumor necrosis factor in rodent endotoxemia: improved survival and organ function. J Infect Dis 1995; 171:1528–1538
- 16. Aiura K, Gelfand JA, Burke JF, et al. Interleukin-1 (IL-1) receptor antagonist prevents *Staphylococcus epidermidis* induced hypotension and reduces circulating levels of tumor necrosis factor and IL-1 beta in rabbits. Infect Immun 1993; 61:3342–3350
- 17. Opal SM, Fisher CF, Dhainaut J-F, et al. Confirmatory interleukin-1 receptor antagonist trial in severe sepsis: a phase III randomized, double-blind, placebo-controlled multicenter trial. Crit Care Med 1997; 25:1115–1124
- 18. Bresnihan B, Alvaro-Gracia JM, Cobby M, et al. Treatment of rheumatoid arthritis with recombinant human interleu-kin-1 receptor antagonist. Arthritis Rheum 1998; 41:2196– 2204
- 19. K.-I. Arai, F. Lee, A. Miyajima, S. Miyatake, N. Arai & T. Yokota: Cytokines: coordinators of immune and inflammatory responses. *Annu Rev Biochem* 59, 783-836 (1991)
- 20. P.S.Crosier & S.C. Clark: Basic biology of the hematopoietic growth factors. *Semin Oncol* 19, 349-61 (1992)
- R. Kuhn, K. Rajewsky & W. Muller: Generation and analysis of interleukin-4 deficient mice. *Science* 254, 707-10 (1991)
- 22. C. Fertin, J.F. Nicolas, P. Gillery, B. Kalis, J. Banchereau & F.X. Maquart: Interleukin-4 stimulates collagen synthesis by normal and scleroderma fibroblasts in dermal equivalents. *Cell Mol Biol* 37, 823-9 (1991)
- 23. C.A. Feghali, K.L. Bost, D.W. Boulware & L.S. Levy: Human recombinant interleukin-4 induces proliferation and interleukin-6 production by cultured human skin fibroblasts. *Clin Immunol Immunopathol* 63, 182-7 (1992)
- 24. P. Miossec: Interleukin 4. A potential antiinflammatory agent. *Rev Rheum* 60, 119-24 (1993)
- 25. Brown MA, Hural J. Functions of IL-4 and control of its expression. Crit Rev Immunol 1997; 17:1–32
- 26. Wang P, Wu P, Siegel ML, et al. Interleukin (IL)-10 inhibits nuclear factor kB (NF-kB) activation in human monocytes: IL-10 and IL-4 suppress cytokine synthesis by different mechanisms. J Biol Chem 1995:9558–9563

- 27. te Velde AA, Huijbens RJF, de Vries JE, et al. Interleukin-4 (IL-4) inhibits secretion of IL-1b, tumor necrosis factor a, and human IL-6 by human monocytes. Blood 1990; 76: 1392–1397
- Paul WE. Interleukin-4: a prototypic immunoregulatory lymphokine. Blood 1991; 77:1859–1870
- 29. Vannier E, Miller MC, Dinarello CA. Coordinated anti-inflammatory effects of interleukin4: interleukin 4 suppresses interleukin 1 production but up-regulates gene expression and synthesis of interleukin 1 receptor antagonist. Proc Natl Acad Sci USA 1992; 89:4076–4080
- 30. Hart PH, Vitti GF, Burgess DR, et al. Potential anti-inflammatory effects of interleukin 4: suppression of human monocyte tumor necrosis factor alpha, interleukin 1, and prostaglandin E2. Proc Natl Acad Sci USA 1989; 86:3803–3807
- 31. Jain-Vora S, LeVine AM, Chroneos Z, et al. Interleukin-4 enhances pulmonary clearance of *Pseudomonas aeruginosa*. Infect Immun 1998; 66:4229–4236
- 32. Hultgren O, Kopf M, Tarkowski A. *Staphylococcus aureus* induced septic arthritis and septic death is decreased in IL-4-deficient mice: role of IL-4 as promoter for bacterial growth. J Immunol 1998; 160:5082–5087
- Toi M, Harris AL, Bicknell R. Interleukin-4 is a potent mitogen for capillary endothelium. Biochem Biophys Res Commun 1991; 174:1287–1293
- 34. Tepper RI, Coffman RL, Leder P. An eosinophil-dependent mechanism of the antitumor effect of IL-4. Science 1992; 257:548–551
- 35. Toi M, Bicknell R, Harris AL. Inhibition of colon and breast carcinoma cell growth by interleukin-4. Cancer Res 1992; 52:275–279
- 36. T. Hirano: The biology of interleukin-6. *Chem Immunol* 51, 153-80 (1992)
- T. Hirano, T. Taga, T. Matsuda, M. Hibi, S. Suematsu, B. Tang, M. Murakami & T. Kishimoto: Interleukin 6 and its receptor in the immune response and hematopoiesis. *Int J Cell Cloning* 8, 155-66 (1990)
- T. Hirano: Interleukin-6 and its relation to inflammation and disease. *Clin Immunol Immunopathol* 62, S60-5 (1992)
- 39. P.L.J. Tan, S. Farmiloe, S. Yeoman & J.D. Watson: Expression of the interleukin 6 gene

in rheumatoid synovial fibroblasts. J Rheumatol 17, 1608-12 (1990)

- C.A. Feghali, K.L. Bost, D.W. Boulware & L.S. Levy: Mechanisms of pathogenesis in scleroderma. I. Overproduction of IL-6 by fibroblasts cultured from affected skin sites of patients with scleroderma. *J Rheumatol* 19, 1207-11 (1992)
- Barton BE. IL-6: insights into novel biological activities. Clin Immunol Immunopathol 1997; 85:16–20
- 42. Barton BE, Shortall J, Jackson JV. Interleukins 6 and 11 protect mice from mortality in a staphylococcal enterotoxininduced toxic shock model. Infect Immun 1996; 64:714–718
- 43. Libert C, Takahashi N, Cauwels A, et al. Response of interleukin-6-deficient mice to tumor necrosis factor-induced metabolic changes and lethality. Eur J Immunol 1994; 24:2237–2242
- 44. Xing Z, Gauldie J, Cox G, et al. IL-6 is an anti-inflammatory cytokine required for controlling local or systemic acute inflammatory responses. J Clin Invest 1998; 101:311–320
- 45. Ruzek MC, Miller AH, Opal SM, et al. Characterization of early cytokine responses in an interleukin-6-dependent pathway of endogenous glucocorticoid induction during murine cytomegalovirus infection. J Exp Med 1997; 185: 1185–1192
- 46. Tilg H, Trehu E, Atkins MB, et al. Interleukin-6 as an anti-inflammatory cytokine: induction of circulating IL-1 receptor antagonist and soluble tumor necrosis factor receptor p55. Blood 1994; 83:113–118
- 47. M.D. Miller & M.S. Krangel: Biology and biochemistry of the chemokines: a family of chemotactic and inflammatory cytokines, *Crit Rev Immunol* 12, 17-46 (1992)
- 48. M.Y. Stoeckle & K.A. Barker: Two burgeoning families of platelet factor 4-related proteins: mediators of the inflammatory response, *New Biologist* 2, 313-23 (1990)
- R.M. Strieter, T.J. Standiford, G.B. Huffnagle, L.M. Colletti, N.W. Lukacs & S.L. Kunke: "The Good, the Bad, and the Ugly." The role of chemokines in models of human disease. J Immunol 156, 3583-86 (1996)
- 50. M. Baggiolini & I. Clark-Lewis: Interleukin-8, a chemotactic and inflammatory cytokine. *FEBS Lett* 307, 97-101 (1992)

- 51. J. Van Damme: Interleukin-8 and related chemotactic cytokines. In: The Cytokine Handbook. Ed: Thomson A., Academic Press, San Diego, CA (1994)
- 52. M. Seitz, B. Dewald, M. Ceska, N. Gerber & M. Baggiolini: Interleukin-8 in inflammatory rheumatic diseases: synovial fluid levels, relation to rheumatoid factors, production by mononuclear cells, and effects of gold sodium thiomalate and methotrexate. *Rheumatol Int* 12, 159-64 (1992)
- 53. Y.R. Mahida, M. Ceska, F. Effenberger, L. Kurlak, I. Lindley & C.J. Hawkey: Enhanced synthesis of neutrophil-activating peptide-1/ Role of cytokines in inflammation 23 interleukin-8 in active ulcerative colitis. *Clin Sci* 82, 273-5 (1992)
- 54. S.L. Kunkel, N. Lukacs, T. Kasama & R.M. Strieter: The role of chemokines in inflammatory joint disease. *J Leukoc Biol* 58, 6-12 (1996)
- 55. Van Snick, A. Goethals, J.C. Renauld, E. Van Roost, C. Uyttenhove, M.R. Rubira, R.L. Moritz & R.J. Simpson: Cloning and characterization of a cDNA for a new mouse T cell growth factor (P40). *J Exp Med* 169, 363-8 (1989)
- 56. Y.C. Yang, S. Ricciardi, A. Ciarletta, J. Calvetti, K. Kelleher & S.C. Clark: Expression cloning of cDNA encoding a novel human hematopoietic growth factor: human homologue of murine T-cell growth factor P40. *Blood* 74, 1880-4 (1989).
- M.C. Cohen & S. Cohen: Cytokine Function. A study in biologic diversity. *Am J Clin Pathol* 105, 589-98 (1996)
- W.F. Chen & A. Zlotnik: IL-10: a novel cytotoxic T cell differentiation factor. J Immunol 147, 528-34 (1991)
- 59. R. de Waal Malefyt, H. Yssel, M.G. Roncarolo, H. Spits & J.E. de Vries: Interleukin-10. *Curr Opin Immunol* 4, 314-20 (1992)
- T.R. Mosmann & K.W. Moore: The role of IL-10 in crossregulation of TH1 and TH2 responses. *Immunol Today* 12, A49-53 (1991)
- 61. K.W. Moore, P. Vieira, D.F. Fiorentino, M.L. Troustine, T.A. Khan & T.R. Mosmann: Homology of cytokine synthesis inhibitory factor (IL10) to the Epstein-Barr virus gene BCRF1. *Science* 248, 1230-4 (1990)

- 62. T.R. Mosmann: Interleukin-10. In: The Cytokine Handbook. Ed: Thomson A., Academic Press, San Diego, CA (1994)
- 63. Lalani I, Bhoi K, Ahmed AF. Interleukin-10: biology, role in inflammation and autoimmunity. Ann Allergy 1997; 79:469–483
- Howard M, O'Garra A. Biological properties of interleukin-10. Immunol Today 1992; 13:198–200
- Opal SM, Wherry JC, Grint P. Interleukin-10: potential benefits and possible risks in clinical infectious diseases. Clin Infect Dis 1998; 27:1497–1507
- 66. Brandtzaeg P, Osnes L, Ovstebo R, et al. Net inflammatory capacity of human septic shock plasma evaluated by a monocyte-based target cell assay: identification of interleukin- 10 as a major functional deactivator of human monocytes. J Exp Med 1996; 184:51–60
- 67. Clarke CJP, Hales A, Hunt A, et al. IL-10 mediated suppression of TNF-a production is independent of its ability to inhibit NF-kB activity. Eur J Immunol 1998; 28:1719–1726
- 68. Ge´ rard D, Bryns C, Marchant A, et al. Interleukin 10 reduces the release of tumor necrosis factor and prevents lethality in experimental endotoxemia. J Exp Med 1993; 177:547–550
- 69. Marchant A, Bruyns C, Vandenabeele P, et al. Interleukin- 10 controls interferon-g and tumor necrosis factor production during experimental endotoxemia. Eur J Immunol 1994; 24:1167–1171
- 70. Dickensheets HL, Freeman SL, Smith MF, et al. Interleukin- 10 upregulates tumor necrosis factor receptor type II (p75) gene expression in endotoxin-stimulated human monocytes. Blood 1997; 90:4162–4171
- 71. Joyce DA, Gibbons DP, Geen P, et al. Two inhibitors of pro-inflammatory cytokine release, interleukin-10 and interleukin- 4, have contrasting effects on release of soluble p75 tumor necrosis factor receptor by cultured monocytes. Eur J Immunol 1994; 24:2699– 2705
- 72. Marchant A, Deviere J, Byl B, et al. Interleukin-10 production during septicaemia. Lancet 1994; 343:707–708
- 73. Van der Poll T, de Waal Malefyt R, Coyle SM, et al. Anti-inflammatory cytokine responses during clinical sepsis and experimental endotoxemia: sequential measurements of plasma soluble interleukin

(IL)-1 receptor type II, IL-10, and IL-13. J Infect Dis 1997; 175:118–122

- 74. Lehmann AK, Halstensen A, Sornes S, et al. High levels of interleukin-10 in serum are associated with fatality in meningococcal
- disease. Infect Immun 1995; 63:2109–211
- 75. Westendorp RGJ, Langermans JAM, Hurizinga TWJ, et al. Genetic influence on cytokine production in fatal meningococcal disease. Lancet 1997; 349:170–173
- Van Dissel JT, van Langevelde P, Westendorp RGJ, et al. Anti-inflammatory cytokine profile and mortality in febrile patients. Lancet 1998; 351:950–953
- 77. Pajkart D, Camoglio L, Tiel-van Buul MCM, et al. Attenuation of pro-inflammatory response by recombinant human IL-10 in human endotoxemia: the effect of timing of rhIL-10 administration. J Immunol 1997; 158:3971–3977
- 78. Dai W, Kohler G, Brombacher F. Both innate and acquired immunity to *Listeria monocytogenes* infections are increased in IL-10-deficient mice. J Immunol 1997; 158:2259–2267
- Van der Poll T, Marchant A, Koeogh CF, et al. Interleukin- 10 impairs host defense in murine pneumococcal pneumonia. J Infect Dis 1996; 174:994–1000
- Greenberger MJ, Strieter RM, Kunkel SL, et al. Neutralization of IL-10 increases survival in a murine model of Klebsiella pneumonia. J Immunol 1995; 155:722–729
- 81. Kuhn R, Lo" hler J, Rennick D, et al. Interleukin-10-deficient mice develop chronic enterocolitis. Cell 1993; 75:263–274
- H. Baumann & P. Schendel: Interleukin-11 regulates the hepatic expression of the same plasma protein genes as interleukin-6. *J Biol chem* 266, 20424-27 (1991)
- 83. Du XX, Williams DA. Interleukin-11: a multifunctional growth factor derived from the hematopoietic micro-environment. Blood 1994; 83:2023–2030
- Neddermann P, Graiziani R, Ciliberto G, et al. Functional expression of soluble interleukin-11 (IL-11) receptor a andstoichiometry of *in vitro* IL-11 receptor complexes with GP130. J Biol Chem 1996; 271:30986–30991
- 85. Tepler I, Elias L, Smith JW, et al. A randomized-placebocontrolled trial of recombinant human interleukin-11 in cancer

patients with severe thrombocytopenia due to chemotherapy. Blood 1996; 87:3607–3614

- Trepicchio WL, Wang L, Bozza N, et al. Interleukin-11 regulates macrophage effector function through the inhibition of nuclear factor-kB. J Immunol 1997; 159:5661–5669
- Hill GR, Cooke KR, Teshima T, et al. Interleukin-11 promotes T cell polarization and prevents acute graft-vs-host disease after allogeneic bone marrow transplantation. J Clin Invest 1998; 201:115–123
- Hermann JA, Hall MA, Maini RN, et al. Important immunoregulatory role of interleukin-11 in the inflammatory process in rheumatoid arthritis. Arthritis Rheum 1998; 41:1388–1397
- 89. M.K. Gately, A.G. Wolitzky, P.M. Quinn & R.Chizzonite: Regulation of human cytolytic lymphocyte responses by interleukin-12. *Cell Immunol* 143, 127-42 (1992)
- 90. M. Kiniwa, M. Gately, U. Gubler, R. Chizzonite, C. Fargeas & G. Delespesse: Recombinant interleukin-12 suppresses the synthesis of immunoglobulin E by interleukin-4 stimulated human lymphocytes. *J Clin Invest* 90, 262-6 (1992)
- 91. P. Scott: IL-12: initiation cytokine for cellmediated immunity. *Science* 260, 496-7 (1993)
- 92. J. Punnonen, G. Aversa, B.G. Cocks, A.N. McKenzie, S. Menon, G. Zurawski, R. de Waal Malefyt & J.E. de Vries JE: Interleukin 13 induces interleukin 4-independent IgG4 and IgE synthesis and CD23 expression by human B cells. *Proc Natl Acad Sci USA* 90, 3730-4 (1993)
- 93. De Waal Malefyt R, Figdor CG, Huijbens R, et al. Effects of IL-13 on phenotype, cytokine production, and cytotoxic function of human monocytes. J Immunol 1993; 151:6370–6381
- 94. Zurawski G, de Vries JE. Interleukin 13, an interleukin-4-like cytokine that acts on monocytes and B cells, but not on T cells. Immunol Today 1994; 15:19–26
- 95. Callard RE, Matthews DJ, Hibbert L. IL-4 and IL-13 receptors: are they one and the same? Immunol Today 1996; 17:108–110
- 96. Mijatovic T, Kruys V, Caput D, et al. Interleukin-4 and -13 inhibit tumor necrosis factor-a mRNA translational activation in lipopolysaccharide-induced mouse macrophages. J Biol Chem 1997; 272:14394– 14398

- 97. Di Santo E, Meazza C, Sironi M, et al. IL-13 inhibits TNF production but potentiates that of IL-6 *in vivo* and *ex vivo* in mice. J Immunol 1997: 159:379–382
- 98. Muchamuel T, Menon S, Pisacane P, et al. IL-13 protects mice from lipopolysaccharideinduced lethal endotoxemia: correlation with down-modulation of TNF-a, IFN-g, and IL-12 production. J Immunol 1997; 158:2898–2903
- 99. Mulligan MS, Warner RL, Foreback JL, et al. Protective effects of IL- 4, IL-10, IL-12, and IL-13 in IgG immune complex-induced lung injury: role of endogenous IL-12. J Immunol 1997; 159:3483–3489
- 100 . Lentsch AB, Czermak BJ, Bless NM, et al. NF-kB activation during IgG immune complex-induced lung injury: requirements for TNF-a and IL-1b but not complement. Am J Pathol 1998; 152:1327–1336
- 101 .Lentsch AB, Czermak BJ, Jordan JA, et al. Regulation of acute lung inflammatory injury by endogenous IL-13. J Immunol 1999; 162:1071–1076
- 102 . Zurawski G, de Vries JE. Interleukin 13, an interleukin-4-like cytokine that acts on monocytes and B cells, but not on T cells. Immunol Today 1994; 15:19–26
- 103 . McKenzie AN, Li X, Largaespada DA, et al. Structural comparison and chromosomal localization of the human and mouse IL-13 genes. J Immunol 1993; 150:5436–5444
- 104 . Callard RE, Matthews DJ, Hibbert L. IL-4 and IL-13 receptors: are they one and the same? Immunol Today 1996; 17:108–110
- 105. Vannier E, Miller MC, Dinarello CA. Coordinated anti-inflammatory effects of interleukin4: interleukin 4 suppresses interleukin 1 production but up-regulates gene expression and synthesis of interleukin 1 receptor antagonist. Proc Natl Acad Sci USA 1992; 89:4076–4080
- 106 .Hart PH, Vitti GF, Burgess DR, et al. Potential anti-inflammatory effects of interleukin 4: suppression of human monocyte tumor necrosis factor alpha, interleukin 1, and prostaglandin E2. Proc Natl Acad Sci USA 1989; 86:3803–3807
- 107 . Jain-Vora S, LeVine AM, Chroneos Z, et al. Interleukin-4 enhances pulmonary clearance of *Pseudomonas aeruginosa*. Infect Immun 1998; 66:4229–4236
- 108. Hultgren O, Kopf M, Tarkowski A. Staphylococcus aureusinduced septic arthritis

and septic death is decreased in IL-4-deficient mice: role of IL-4 as promoter for bacterial growth. J Immunol 1998; 160:5082–5087

- 109 .Toi M, Harris AL, Bicknell R. Interleukin-4 is a potent mitogen for capillary endothelium. Biochem Biophys Res Commun 1991; 174:1287–1293
- 110 . J. Vilcek, T.H. Lee: Tumor necrosis factor. *J Biol Chem* 266, 7313-16 (1991)
- 111 . R.A. Smith & C.Baglioni: The active form of tumor necrosis factor is a trimer. J Biol Chem 262, 6951-4(1987)
- 112. B.B. Aggarwal: Tumor Necrosis Factor. In: Human cytokines. Eds: Aggarwal B.B., Gutterman J.U., Blackwell Scientific Publications, Boston, MA (1992)
- 113. B. Beutler & A. Cerami: The common mediator of shock, cachexia, and tumor necrosis. *Adv Immunol* 42, 213-31 (1988)
- 114 . N.L. Paul & N.H. Ruddle: Lymphotoxin. Ann Rev Immunol 6, 407-38 (1988)
- 115 . T. Miethke, H. Gaus, C. Wahl, K. Heeg & H. Wagner: T-cell-dependent shock induced by a bacterial superantigen. *Chem Immunol* 55, 172-84 (1992)
- 116. T. Miethke, C. Wahl, K. Heeg, B. Echtenacher, P.H. Krammer & H. Wagner: T cell-mediated lethal shock triggered in mice by the superantigen staphylococcal enterotoxin B: critical role of tumor necrosis factor. *J Exp Med* 175, 91-8 (1992)
- 117. E.A. Garcia-Zepeda, M.E. Rothenberg, R.T. Ownbey, J. Celestin, P. Leder & A.D. Luster: Human eotaxin is a specific chemoattractant for eosinophil cells and provides a new mechanism to explain tissue eosinophilia. *Nature Med* 2, 449-56 (1996)
- 118. D. Metcalf: The colony stimulating factors. Discovery, development, and clinical applications. *Cancer* 65, 2185-95 (1990)
- 119. S. Nagata: Granulocyte colony stimulating factor and its receptor. In: The Cytokine Handbook. Ed: Thomson A., Academic Press, San Diego, CA (1994)
- 120 . W.A. Border & E. Ruoslahti: Transforming growth factor- $\beta$  in disease: the dark side of tissue repair. *J Clin Invest* 90, 1-7 (1992)
- 121. M.B. Sporn, A.B. Roberts, L.M. Wakefield & R.K. Assoian RK: Transforming growth factor-β : biological function and chemical structure. *Science* 233, 532-4 (1986)
- 122 . W.O. Cooper, R.A. Fava, C.A. Gates, M.A. Cremer & A.S. Townes: Acceleration of onset

of collagen-induced arthritis by intra-articular injection of tumour necrosis factor or transforming growth factor-beta. *Clin Exp Immunol* 89, 244-50 (1992)

- 123. E.F. Wheelock: Interferon-like virusinhibitor induced in human leukocytes by phytohemagglutinin. *Science* 149, 310-11 (1965)
- 124. E. De Maeyer & J. De Maeyer-Guignard: Interferons. In: The Cytokine Handbook. Ed: Thomson A., Academic Press, San Diego, CA (1994)
- 125 . H. Heremans & A. Billiau: The potential role of interferons and interferon antagonists in inflammatory disease. *Drugs* 38, 957-72 (1989)
- 126 . C.O. Jacob, P.H. van der Meide & H.O. McDevitt: In vivo treatment of (NZB x NZW)F1 lupus-like nephritis with monoclonal antibody to  $\gamma$ - interferon. J Exp Med 166, 798-803 (1987)
- 127 . I.L. Campbell, L. Oxbrow, M. Koulmanda & L.C. Harrison: IFN-γ induces islet cell MHC antigens and enhances autoimmune, streptozotocin-induced diabetes in the mouse. *J Immunol* 140, 1111-6 (1988)
- 128. N. Sarvetnick, D. Liggitt, S.L. Pitts, S.E. Hansen & T.A. Stewart: Insulin-dependent diabetes mellitus induced in transgenic mice by ectopic expression of class II MHC and interferon-gamma. *Cell* 52, 773-82 (1988)
- 129. N.J. Mauritz, R. Holdmdahl, R. Jonsson, P.H. van der Meide, A. Scheynius & L. Klareskog: Treatment with gamma-interferon triggers the onset of collagen arthritis in mice. *Arthritis Rheum* 31, 1297-1304 (1988)
- 130. G.E. Grau, H. Heremans, P.F. Piguet, P. Pointaire, P.H. Lambert, A. Billiau & P. Vassalli: Monoclonal antibody against interferon can prevent experimental cerebral malaria associated and its overproduction of tumor necrosis factor. Proc Natl Acad Sci USA 86, 55724 (1989)
- 131. D.K. Dalton, S. Pitts-Meek, S. Keshav, I.S. Figari, A. Bradley & T.A. Stewart: Multiple defects of immune cell function in mice with disrupted interferon-gamma genes. *Science* 259, 1739-42 (1993)
- 132. H. Okamura, H. Tsutsui, T. Komatsu, M. Yutsudo, A. Hakura, T. Tanimoto, K. Torigoe, T. Okura, Y. Nukada, K. Hattori, K. Akita, M. Namba, F. Tanabe, K. Konishi, S. Fukuda, & M. Kurimoto: Cloning of a new cytokine that

induces IFN-γ production by T cells. *Science* 378, 88-91 (1995)

- 133. S. Ushio, M. Namba, T. Okura, K. Hattori, Y. Nukada, K. Akita, F. Tanabe, K. Konishi, M. Micallef, M. Fuji, K. Torigoe, T. Tanimoto, S. Fukuda, M. Ikeda, H. Okamura, & M. Kurimoto: Cloning of the cDNA for human IFN- -inducing factor, expression in *Escherichia coli*, and studies on the biologic activities of the protein. *J Immunol* 156, 4274-9 (1996)
- 134 . Van der Poll T, Jansen J, van Leenen D, et al. Release of soluble receptors for tumor necrosis factor in clinical sepsis and experimental endotoxemia. J Infect Dis 1993; 168:955-960
- 135. Ertel W, Scholl FA, Galatti H, et al. Increased release of soluble tumor necrosis factor receptors into blood during clinical sepsis. Arch Surg 1994; 129:1330–1337
- 136. Van Deuren M, Frieling TM, van der ven-Jongekrijg J, et al. Plasma patterns of tumor necrosis factor-a and TNF soluble receptors during acute meningococcal infections and the effect of plasma exchange. Clin Infect Dis 1998; 26:918–923
- 137 . Mohler KM, Torrance DS, Smith CA, et al. Soluble tumor necrosis factor (TNF) receptors are effective therapeutic agents in lethal endotoxemia and function simultaneously as both TNF carriers and TNF antagonists. J Immunol 1993; 151:1548–1561
- 138 . Evans TJ, Moyes D, Carpenter A, et al. Protective effect of 55- but not 75-kD soluble tumor necrosis factor receptorimmunoglobulin G fusion proteins in an animals model of Gram-negative sepsis. J Exp Med 1994; 180:2173–2179
- 139. Van Zee KJ, Kohno T, Fisher E, et al. Tumor necrosis factor soluble receptors circulate during experimental and clinical inflammation and can protect against excessive tumor necrosis factora *in vitro* and *in vivo*. Proc Natl Acad Sci USA 1992; 89:4845–4849
- 140 . Aizawa Y, Akita K, Taniai M, et al. Cloning and expression of interleukin-18 binding protein. FEBS Lett 1999; 445:338–342
- 141 . Yoshimoto T, Takeda K, Tanaka T, et al. IL-12 up-regulates IL-18 receptor expression on T cells, Th1 cells, and B cells: synergism with IL-18 for IFN-g production. J Immunol 1998; 161:3400–3407

- 142 . Thomassen E, Bird TA, Renshaw BR, et al. Binding of interleukin-18 to the interleukin-1 receptor homologous receptor IL-1Rrpl leads to activation of signaling pathways similar to those used by interleukin-1. J Interferon
  - Cytokine Res 1998; 18:1077–1088
- 143. Hoshino K, Tsutsui H, Kawai T, et al. Cutting edge: generation of IL-18 receptordeficient mice; evidence for IL-1 receptorrelated protein as an essential IL-18 binding receptor. J Immunol 1999; 62:5041–5044
- 144. Kanakaraj P, Ngo K, Wu Y, et al. Defective interleukin (IL)-18-mediated natural killer and T helper cell type 1 responses in IL-1 receptor-associated kinase (IRAK)-deficient mice. J Exp Med 1999; 189:1129–1138
- 145 . Spriggs MK. One step ahead of the game: viral immunomodulatory molecules. Annu Rev Immunol 1996; 14:101–130
- 146 . Ploegh HL. Viral strategies of immune evasion. Science 1998; 280:248–253
- 147 . Moore KW, Vieira P, Fiorentino DF, et al. Homology of cytokine synthesis inhibitory factor (IL-10) to Epstein-Barr virus gene *BCRF1*. Science 1990; 248:1230–1234
- 148. Wilson M, Seymour R, Henderson B. Bacterial perturbation of cytokine networks. Infect Immun 1998; 66:2401–2409
- 149 . Goldie AS, Fearon KCHM, Ross JA, et al. Natural cytokine antagonists and endogenous anti-endotoxin core antibodies in sepsis syndrome. JAMA 1995; 274:172–177
- 150. Girardin E, Roux-Lomberd P, Grau GE, et al. Imbalance between tumor necrosis factor-a and soluble TNF receptor concentration in severe meningococcaemia. Immunology 1992; 76:20–23
- 151 . Moreland LW, Baumgartner SW, Schiff MH, et al. Treatment of rheumatoid arthritis with a recombinant human tumor necrosis factor receptor (p75)-Fc fusion protein. N Engl J Med 1997; 337:141–147.
- 152. T. Kishimoto, S. Akira & T. Taga: Interleukin-6 and its receptor: a paradigm for cytokines. *Science* 258, 593-7 (1992)
- 153. W.C. Greene: The human interleukin-2 receptor. *Annu Rev Immunol* 4, 69-95 (1986)
- 154. K.A. Smith: The two-chain structure of highaffinity IL-2 receptors. *Immunol Today* 8, 11-13 (1987)
- 155 . T. Taga, M. Hibi, M. Murakami, M. Saito, H. Yawata, M. Narazaki, Y. Hirata, T. Sugita, K. Yasukawa, T. Hirano & T. Kishimoto:

Interleukin-6 receptor and signals. *Chem Immunol* 51, 181-204 (1992)

- 156 . J.F. Bazan: Structural design and molecular evolution of a cytokine receptor superfamily. *Proc Natl Acad Sci USA* 87, 6934-8 (1990)
- 157. A. Oehm, I. Behrmann, W. Falk, M. Pawlita, G. Maier, C. Klas, M. Li-Weber, S. Richards, J. Dhein, B.C. Trauth, H. Ponstingl & P.H. Krammer: Purification and molecular cloning of the APO-1 cell surface antigen, a member of the tumor necrosis factor/nerve growth factor receptor superfamily: sequence identity with the Fas antigen. J Biol Chem 267, 10709-15 (1992)
- 158. J. Banchereau, F. Bazan, D. Blanchard, F. Briere, J.P. Galizzi, C. van Kooten, Y.J. Liu, F. Rousset, & S. Saeland: The CD40 antigen and its ligand. *Annu Rev Immunol* 12, 881-922 (1994)
- 159 . L.A. Tartaglia & D.V. Goeddel: Two TNF receptors. *Immunol Today* 13, 151-3 (1992)
- 160 . K. Wiegmann, S. Schutze, E. Kampen, A. Himmler, T. Machleidt & M. Kronke: Human 55-kDa receptor for tumor necrosis factor

coupled to signal transduction cascades. *J Biol Chem* 267, 17997-8001 (1992)

- 161. T.N.C. Wells, C.A. Power, M. Lusti-Narasimhan, A.J. Hoogewerf, R.M. Cooke, C. Chung, M.C. Peitsch & A.E.I. Proudfoot:
- Selectivity and antagonism of chemokine receptors. J Leukoc Biol 59, 53-60 (1996)
- 162. C.J. Raport, V.L. Schweickart, D. Chantry, R.L. Eddy Jr., T.B. Shows, R. Godiska & P.W. Gray: New members of the chemokine receptor gene family. *J Leuko. Biol* 59, 18-23 (1996)
- 163 . P. Bates: Chemokine receptors and HIV-1: an attractive pair? *Cell* 86, 1-3 (1996)
- 164 . M. Dean, M. Carrington, C. Winkler, G.A. Huttley, M.W. Smith, R. Allidmets, J.I. Goedert, S.P. Buchbinder, E. Vittinghoff, E. Gomperts, S. Donfield, D. Vlahov, R. Kaslow, A. Saah, C. Rinaldo & R. Detels: Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the *CKR5* structural gene. *Science* 273, 1856-62 (1996)